## Appendix 1.

## **VSX1 Primers and conditions.**

# VSX1 PCR conditions

# **PCR CONDITIONS:**

Used FastStart Taq enzyme kit from Roche, with 3.5mM final concentration of MgCl<sub>2</sub>.

Primer sequences as follows:

Exon1 VSX1\_Ex1aF: 5'-TGCTTGCTAAGGAACCATGA-3'

VSX1\_Ex1cR: 5'-CTCAGAGCCTAGGGGACAGG-3'

Exon2 VSX1\_Ex2F: 5'-AAATCCAGGAAATAGAGGGGA-3'

VSX1\_Ex2R: 5'-AGATGCAGGTGCCATAAACC-3'

Exon3 VSX1\_Ex3F: 5'-CTGTGTGTTTTTGGGGTCCTT-3'

VSX1\_Ex3R: 5'-GTGGTATCTTTGGAGCGGAG-3'

Exon4 VSX1\_Ex4F: 5'-TCTCCTGCCTCCAACCAG-3'

VSX1\_Ex4R: 5'-CTCCTACAACACCTCGAGCC-3'

Exon5 VSX1\_Ex5aF2: 5'-CAATTCTAGTGGGATTTAGAGAACA-3'

VSX1\_Ex5bR: 5'-CCTTTGACAGTGGGACCTGT-3'

Exon6 VSX1\_Ex6F: 5'-ATCTCTTTGCATTCTCAGAGGGTGAGA-3'

VSX1\_Ex6R: 5'-TATTCCTGCTGCATGGGTCCATTTGT-3'

Exon7 VSX1\_Ex7aF2: 5'- CAGTCCCACCTTTCCCTCTCT-3'

VSX1\_Ex7cR2: 5'- CTGCCCTAACACTCAGGGTC-3'

## PCR cycle as follows:

	temperature	time
initial denaturation	96°C	5min
35x -denaturation	94°C	30secs
-annealing	*°C	1min
-extension	72°C	1min
final extension	72°C	10min

\*56°C for exons 1, 2, 3, 7

58°C for exon 4

61°C for exon 5

60°C for exon 6

5uL of product was visualised on 1.5% agarose gel, and the remaining 20ul cleaned with HighPure PCR purification kit from Roche.

## VSX1- Control screen for c.173C>T, p.Pro58Leu

#### **HRM CONDITIONS:**

- Used SensiMix HRM kit from Quantace, with 3mM final concentration of MgCl<sub>2</sub>.
- Primers were designed using NCBI primer-BLAST. Sequences as follows:

**VSX1\_Ex1aF2:** 5'-TTGCTAAGGAACCATGACCG-3'

**VSX1\_Ex1aR2:** 5'-ACAGAGGAGGCCGAGTCC-3'

HRM cycle as follows:

	degC	time
initial denaturation	95	10min
50x -denaturation	95	10secs
-annealing	58	15secs
-extension	72	20secs
HRM step	87-97	N/A

#### **PCR CONDITIONS:**

Used FastStart Taq enzyme from Roche, with 3.5mM final concentration of MgCl<sub>2</sub>.

Primer sequences as follows:

VSX1\_Ex1aF: 5'-TGCTTGCTAAGGAACCATGA-3' - VSX1\_Ex1cR: 5'-CTCAGAGCCTAGGGGACAGG-3'

PCR cycle as follows:

	degC	time
initial denaturation	96	5min
35x -denaturation	94	30secs
-annealing	56	1min
-extension	72	1min
final extension	72	10min

• 5uL of product was visualised on 1.5% agarose gel, and the remaining 20ul cleaned with HighPure PCR purification kit from Roche.

# VSX1- Control screen for c.731A>G, p.His244Arg

## **HRM CONDITIONS:**

- Used High Resolution Melting Master kit from Roche, with 2mM final concentration of MgCl<sub>2</sub>.
- Primers were designed using NCBI primer-BLAST. Sequences as follows:

VSX1\_Ex4F 5'-TCTCCTGCCTCCAACCAG-3'

VSX1\_Ex4R 5'-CTCCTACAACACCTCGAGCC-3'

HRM cycle as follows:

	degC	time
initial denaturation	95	10min
50x -denaturation	95	10secs

-annealing	61	15secs
-extension	72	20secs
HRM step	87-97	N/A

### **PCR CONDITIONS:**

- Used AmpliTaq enzyme from Applied Biosystems, with 1.5mM final concentration of MgCl<sub>2</sub>.
- Primer sequences as above.
- PCR cycle as follows:

	degC	time
initial denaturation	96	5min
35x -denaturation	94	30secs
-annealing	56	1min
-extension	72	1min
final extension	72	10min

• 5uL of product was visualised on 1.5% agarose gel, and the remaining 20ul cleaned with HighPure PCR purification kit from Roche.

# **SEQUENCING CONDITIONS:**

- Used BigDye Terminator from Applied Biosystems.
- Sequenced in both directions, using above PCR primers.
- Sequencing products cleaned by EtOH/NaOAc precipitation, then sent to Allan Wilson Centre (Massey University) for capillary separation.
- Sequencing analysis done using CodonCode Aligner version 3.5.6

#### **REFERENCE SEQUENCES:**

- Genomic: NCBI Reference Sequence NG\_008101.1
- mRNA: NCBI Reference Sequence NM\_014588.4