

Effects of a novel DNA methyltransferase inhibitor Zebularine on human lens epithelial cells

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Purpose: Posterior capsular opacification (PCO) is a common long-term complication of modern cataract surgery. We have shown that Zebularine, an inhibitor of DNA methylation, suppresses transforming growth factor- β (TGF β)-induced lens epithelial cells (LECs)-myofibroblasts transdifferentiation. The purpose of this study is to evaluate the role that Zebularine plays in the inhibition of PCO pathogenesis, including its effect on attachment, migration, and proliferation of LECs in vitro.

Methods: A tetrazolium dye-reduction assay (MTT test) was performed to determine cell proliferation. Cell attachment was assessed by modified MTT test. Migration was determined by the transwell method after incubation of LECs with Zebularine. The effect of Zebularine on DNA-methyltransferase 1 (DNMT1), phospho-p44/42 Map Kinase, and protein kinase B (Akt) were analyzed by western blot.

Results: Zebularine was an effective inhibitor of human LEC proliferation, attachment, and migration in vitro. A Zebularine concentration of 100 μ M accounted for the following: inhibition of cell proliferation of 57.2%, reduction in cell attachment to 29.6%, and inhibition of cell migration of 58.9%. All effects were dose dependent. Zebularine treatment resulted in dose-dependent decreases of DNMT1, phosphorylated p44/42 MAP Kinase, and phosphorylated Akt.

Conclusions: Zebularine is capable of inhibiting the crucial cellular events in PCO pathogenesis in vitro. Zebularine acts through the inhibition of DNMT1, and it consequently down regulation of the expression of proliferative and survival genes that relate to pathogenesis of PCO. These findings suggest that Zebularine may become a therapeutic approach for the prevention of PCO.

The continuous improvements in surgical technique, intraocular lens (IOL) design, and IOL material have significantly reduced the incidence of posterior capsule opacification (PCO) in the past 30 years [1]. However, PCO remains the most common long-term complication of modern cataract surgery [2,3]. Decreased visual acuity induced by PCO is reported to occur in 20% to 40% of patients 2 to 5 years after surgery [3,4]. As a routine treatment for PCO, neodymium: YAG (Nd:YAG) laser capsulotomy is performed successfully in many patients. However, there are complications such as retinal detachment, damage to the intraocular lens, and cystoid macular edema [5,6].

Cataract surgery induces a wound-healing response in the lens, and residual lens epithelial cells (LECs) undergo an epithelial-to-mesenchymal transition (EMT), followed by enhanced proliferation, migration, and collagen deposition. Therefore, postsurgical medical inhibition of LECs EMT, proliferation, and migration is a possible option for preventing PCO.

Epigenetic modifications are post-transcriptional, reversible events that do not target gene sequence, and inhibition of these mechanisms could theoretically be

advantageous in the treatment of fibrosis disease [7,8]. As a consequence, the role of epigenetic regulators like histone deacetylase (HDAC) and DNA methyltransferase (DNMT) inhibitors as a treatment for PCO is under evaluation.

Zebularine is a cytidine analog containing a 2-(1H)-pyrimidinone ring that was originally developed as a cytidine deaminase inhibitor. It acts primarily as a trap for DNMT protein by forming tight covalent complexes between DNMT protein and Zebularine-substituted DNA [9]. In contrast to other DNMT inhibitors, it is quite stable [10,11] and low toxicity [12-14]. Preclinical studies using Zebularine have shown favorable toxicity and stability profiles, making it an attractive candidate for epigenetic treatment of PCO [13].

The purpose of this study was to determine whether Zebularine is capable of inhibiting the crucial cellular events for PCO formation (i.e., human LEC proliferation, attachment, and migration) in vitro.

METHODS

The institutional review board (IRB) of Fudan University Eye and ENT Hospital, Shanghai, China approved our use of cultured human LECs. All procedures conformed to the Declaration of Helsinki for research involving human subjects. The Zebularine we used was a kind gift from Dr. Victor E. Marquez (Laboratory of Medicinal Chemistry, National Cancer Institute, Frederick, MD).

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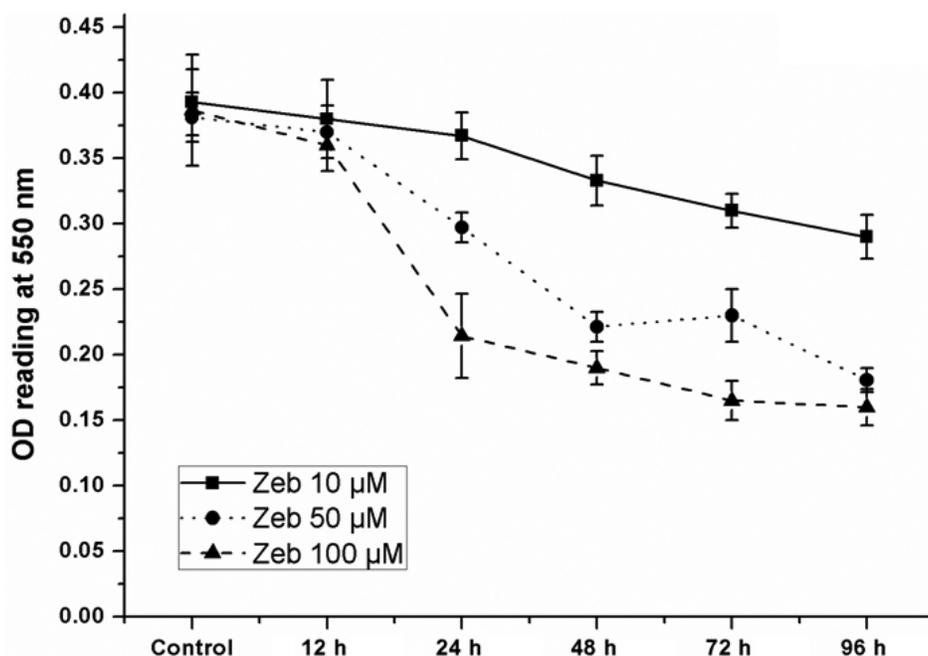


Figure 1. Zebularine inhibited LECs proliferation in a dose- and time-dependent manner. The human LECs were treated with various concentrations of Zebularine and the percentage of surviving cells was determined by MTT assay from 12 to 96 h. A time- and dose-dependent decrease in proliferation was seen with the application of Zebularine (10, 50, and 100 μ M; $p < 0.01$). A Zebularine concentration of 100 μ M accounted for the inhibition of cell proliferation of 57.2% in 96 h.

Cell culture: HLE B-3 cells, an immortalized cell line derived from infant human lens tissue and transformed with adenovirus 12-simian virus (SV40), were obtained from ATCC (Rockville, MD) and cultured in Eagle's minimum essential medium (GIBCO BRL, Grand Island, NY) with 20% fetal bovine serum, 100 units/ml penicillin, and 100 mg/ml streptomycin at 37 °C in a humidified 5% CO₂ atmosphere.

Cell proliferation assay: HLE B-3 cells were seeded in 96-well plates (150 μ l/well at a density of 5×10^3 cells/well in DMEM containing 10% FBS). After 24 h, Zebularine (10, 50, and 100 μ M) was added. After another 12, 24, 48, 72, and 96 h, cells were washed gently twice with PBS, and fresh medium (150 μ l) was added to each well with MTT (3-(4, 5-dimethyl-2-thiazolyl)-2, 5-diphenyl-2H tetrazolium bromide, 5mg/ml, 20 μ l; Sigma, St. Louis, MO). After 4 h of incubation, the supernatants were decanted, and the formazan precipitates were solubilized by the addition of 150 μ l of 100% DMSO (Sigma) and placed on a plate shaker for 10 min. Absorbance at 550 nm was determined on a multi-well plate reader (Benchmark plus™; Bio-Rad, Tokyo, Japan). The number of proliferated cells was proportional to the absorbance of MTT at 550 nm. All experiments were performed at least three times.

Cell attachment assay: The attachment assay was performed by a modified MTT assay using 96-well plates coated with fibronectin. After treatment with Zebularine (10, 50, and 100 μ M) for 48 h, LECs were trypsinized and re-suspended in DMEM with 0.4% FBS. One hundred microliters of cell suspension (10^4 cells) were added to each well and allowed to attach for 5 and 15 min. The cells were washed gently twice with PBS, and fresh medium (150 μ l)

was added to each well with MTT (5 mg/ml, 20 μ l; Sigma). After 4 h of incubation, the supernatants were decanted, and the formazan precipitates were solubilized by the addition of 150 μ l of 100% DMSO (Sigma) and placed on a plate shaker for 10 min. Absorbance at 550 nm was determined on a multi-well plate reader (Benchmark plus™; Bio-Rad). The number of attached cells was proportional to the absorbance of MTT at 550 nm. All experiments were performed at least three times.

Cell migration assay: Migration was measured using Transwell (Costar, Cambridge, MA) assay, as previously described [15]. Briefly, 5×10^4 LECs were seeded in the upper part of a chamber in 24-well plates after treatment with Zebularine (10, 50, and 100 μ M) for 48 h, with inserts coated with fibronectin (2 μ g/cm²). The lower chamber was filled with 0.4% FBS-DMEM containing 20ng/ml recombinant PDGF (R&D Systems Inc., Minneapolis, MN). After 5 h incubation, the inserts were washed three times with PBS, fixed with cold methanol (4 °C) for 10 min, and counterstained with hematoxylin for 20 min. The number of migrated cells was counted by phase-contrast microscopy (200 \times). Three randomly chosen fields were counted per insert. All experiments were performed at least three times.

Western blot assay: Confluent cells grown in 6-well plates in DMEM with 0.4% FBS were lysed, supernatants were collected, and proteins were resolved on Tris-HCl 10% polyacrylamide gels at 120 V. The proteins were transferred to PVDF blotting membrane (Millipore, Bedford, MA). The membranes were probed with antibodies for DNA-methyltransferase 1 (DNMT1; Abcam, Cambridge, MA), phospho-p44/42 MAP Kinase (Thr 202/Thy 204; Cell

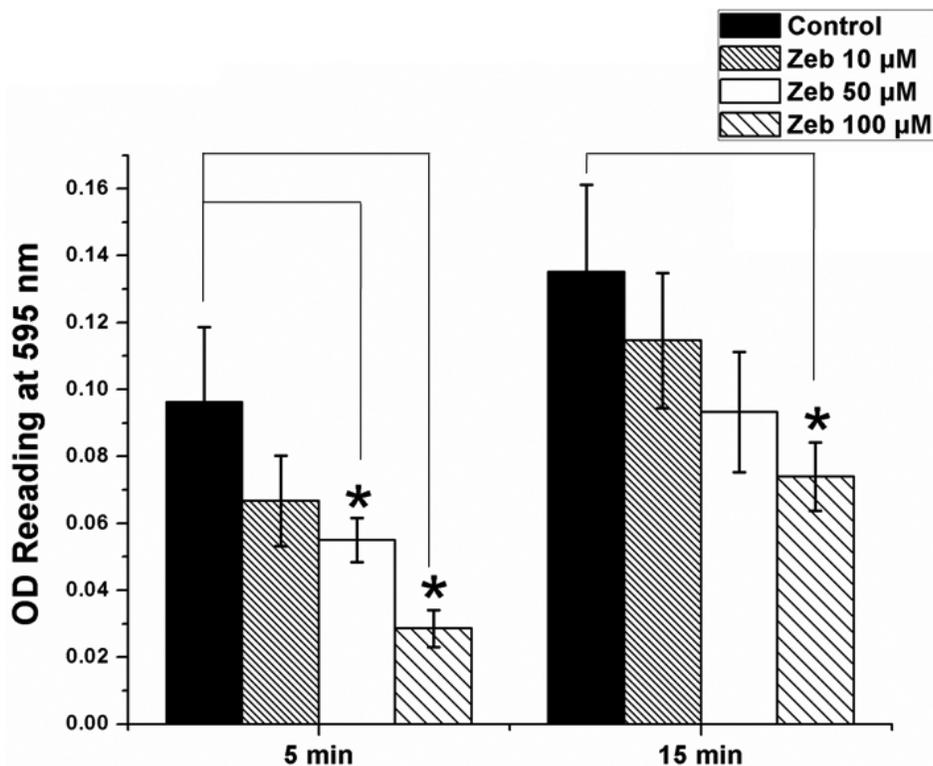


Figure 2. Zebularine decreased LECs attachment in a dose-dependent manner. LECs attachment was measured using modified MTT test. A dose-dependent decrease in attachment was seen with the application of high concentrations of Zebularine (50 µM and 100 µM). LECs attachment was significantly reduced to 29.6% in the early phase (5 min) of cell attachment. * $p < 0.01$, Error bar indicates mean \pm SEM.

Signaling, Danvers, MA), phospho-AKT (Ser 473; Cell Signaling), and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) (Abcam), respectively, all at 1:1,000 dilution. Membranes were washed and incubated with a horseradish peroxidase (HRP)-conjugated secondary antibody (1:3,000; Vector Laboratories, Burlingame, CA) for 30 min at room temperature. Images were developed using ECL chemiluminescence detection solution (Amersham Pharmacia Biotech, Cleveland, OH). All experiments were performed at least three times.

Statistics: The data were analyzed using Student's *t*-test, and a $p < 0.05$ was accepted as significant.

RESULTS

Zebularine inhibited LECs proliferation in a dose- and time-dependent manner: The human LECs were treated with various concentrations of Zebularine, and the percentage of surviving cells was determined by MTT assay from 12 to 96 h. A time- and dose-dependent decrease in proliferation was seen with the application of Zebularine (10, 50, and 100 µM; $p < 0.01$). As shown in Figure 1, 25.6 \pm 4.1% inhibition of cells occurred with 10 µM Zebularine at 96 h, 52.6 \pm 2.3% inhibition with 50 µM Zebularine, and 57.2 \pm 3.3% inhibition with 100 µM Zebularine.

Zebularine decreased LECs attachment: LECs attachment was measured using modified MTT test (Figure 2). After 48 h exposure to Zebularine, no statically significant difference was found in LECs attachment between control and low-dose

Zebularine (10 µM)-treated cells. However, a dose-dependent decrease in attachment was seen with the application of higher concentrations of Zebularine (50 µM and 100 µM; $p < 0.01$). LECs attachment was significantly reduced by 70.4 \pm 5.3% in the early phase (5 min) of cell attachment ($p < 0.01$).

Zebularine inhibited LECs migration: LECs migration to the lower compartment of a transwell was significantly increased in response to PDGF stimulation (Figure 3). PDGF-induced LECs migration was inhibited by Zebularine (50 and 100 µM) in a dose-dependent manner. The maximal inhibition (58.9 \pm 6.1%) was seen at 100 µM Zebularine ($p < 0.01$).

Zebularine inhibited the epigenetic regulator: Because of Zebularine's activity as a DNMT inhibitor in other model systems, its effect on expression of DNMT1 in LECs was examined. As expected, Zebularine treatment was associated with a dose-dependent depletion of DNMT1 in all three doses (10, 50, and 100 µM; Figure 4).

Zebularine inhibited the cellular survival genes: Akt is involved in cellular survival pathways and is active after being phosphorylated at serine 473. In this study, Zebularine inhibited Akt phosphorylation at serine 473 in all three doses. Mitogen-activated protein kinases (MAPKs) are a widely conserved family of serine/threonine protein kinases involved in many cellular programs such as cell proliferation, differentiation, motility, and death. The p44/42 MAPK signaling pathway can be activated in response to a diverse range of extracellular stimuli including mitogens, growth

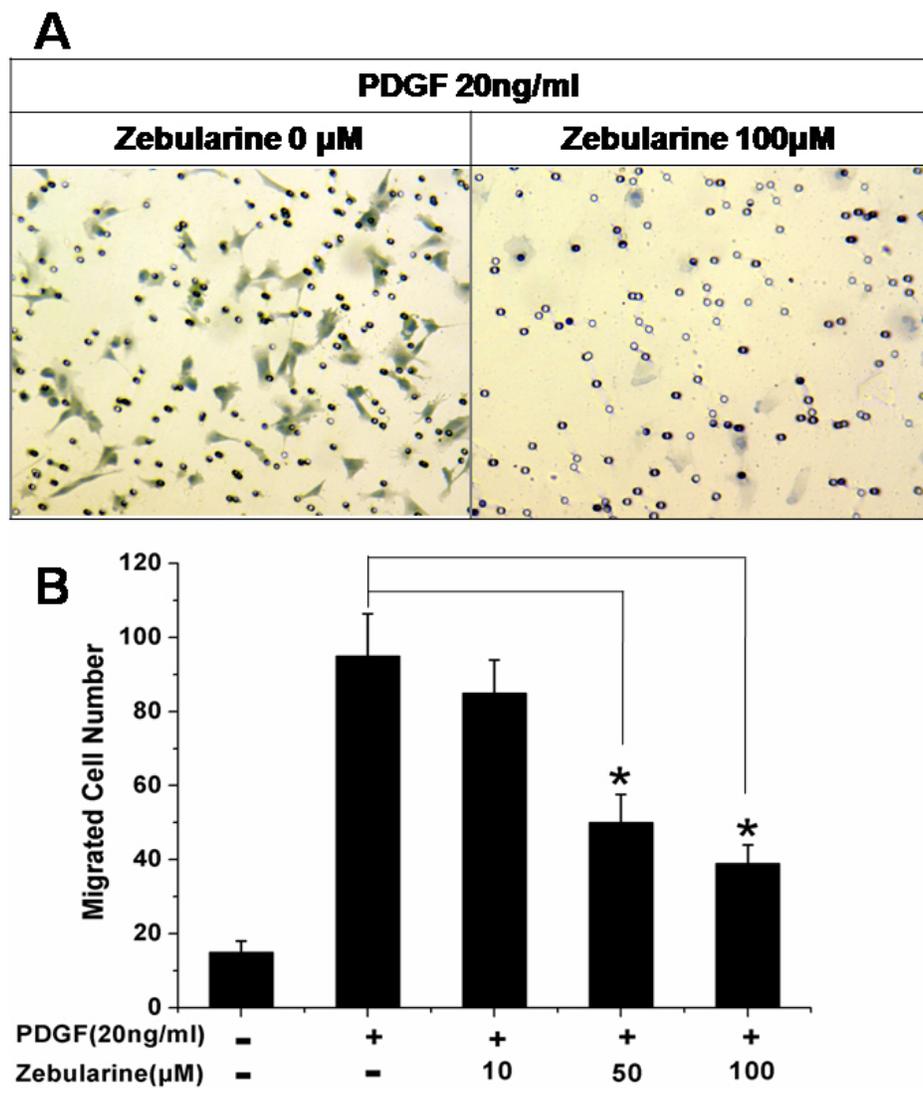


Figure 3. Zebularine inhibited LECs migration in a dose-dependent manner. LECs migration to the lower compartment of a transwell was significantly increased in response to PDGF stimulation. PDGF-induced LECs migration was inhibited by Zebularine (A) in a dose-dependent manner (B). The maximal inhibition (58.9±6.1%) was seen at 100 μ M Zebularine (p<0.01). *p<0.01, Error bar indicates mean±SEM.

factors, and cytokines. In this study, phosphorylated p44/42 MAPK level decreased by 87.5% after treatment with high dose (100 μ M) Zebularine for 48 h (Figure 4).

DISCUSSION

In this study, we found that Zebularine is capable of inhibiting the crucial cellular events in PCO pathogenesis in vitro. After a single exposure to Zebularine added to the cell-culture medium in the presence of serum, HLE-B3 proliferation, attachment, and migration were effectively inhibited.

The initial cellular events in PCO can be described as the migration and attachment of LECs to the posterior capsule surfaces and their following proliferation. In this study, Zebularine effectively inhibited LECs attachment and migration. Inhibition of the initial stages in PCO development may prevent the disease from progressing to the more advanced stages commonly associated with a severe reduction in visual acuity.

The effect of Zebularine on cell migration was reported by Hellebrekers et al. [16]. However, they reported that migration of endothelial cells was not significantly influenced by treatment with Zebularine at concentrations up to 500 μ M. This difference may be due to different type of cells and methods used in different experiments. Hellebrekers et al. [16] used human umbilical vein endothelial cells (HUVEC) and scratched wound assay in response to Zebularine treatment, while we used fetal LECs and transwell assay. Hellebrekers's method more likely showed a wound healing response, while the transwell assay method showed the effect of Zebularine on LECs chemotaxis response. In addition, the pretreatment with Zebularine was 48 h in our experiment, while Hellebrekers et al. [16] pretreated the cells for 72 h.

Proliferation is another important event in the pathogenesis of PCO. Our results demonstrate that Zebularine effectively inhibited LECs proliferation in a time- and dose-dependent manner. Previous studies showed that Zebularine

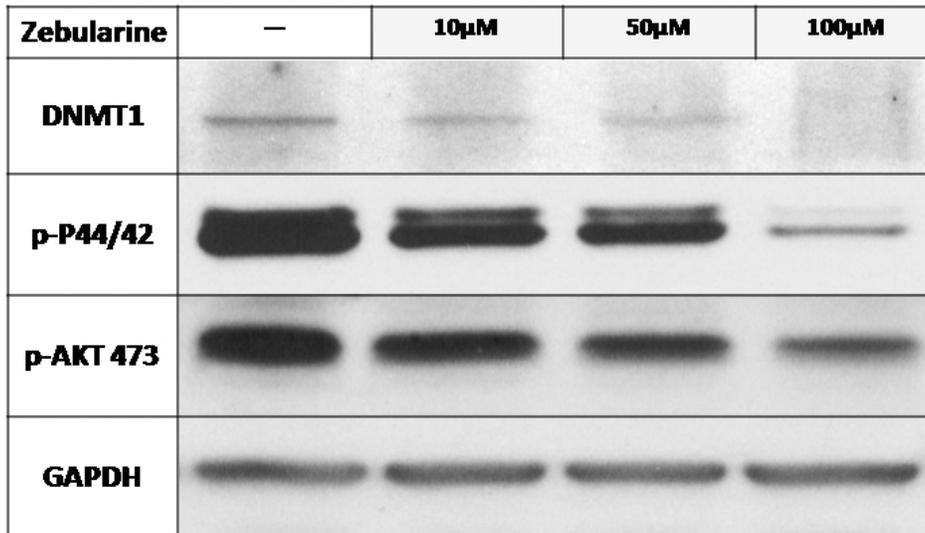


Figure 4. The effect of Zebularine on the expression of methylation related genes and genes involved in the pathogenesis of PCO. Zebularine treatment was associated with a dose-dependent depletion of DNMT1 in all three doses (10, 50, and 100 μ M). Zebularine inhibited the phosphorylation of Akt at serine 473. Phosphorylated p44/42 MAPK level decreased by 87.5% after treatment with high dose (100 μ M) Zebularine in 48 h.

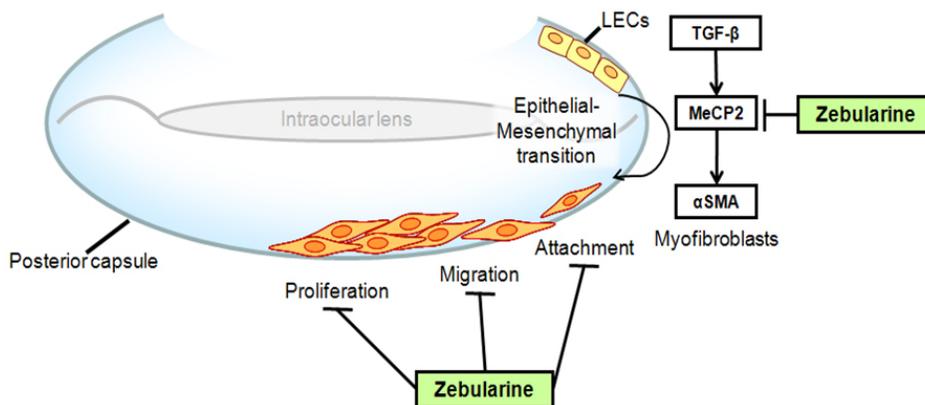


Figure 5. Model for Zebularine inhibiting PCO. We have shown that Zebularine inhibited MeCP2, and furthermore, inhibits TGF β -induced α -SMA expression. In this study, we found that Zebularine is capable of inhibiting the crucial cellular events in PCO pathogenesis, including LECs proliferation, attachment, and migration. Therefore, Zebularine could potentially prevent PCO formation. LECs: lens epithelial cells; IOL: intra ocular lens.

induces apoptotic cell death in many cell lines. Scott et al. [13] analyzed the effects of Zebularine on cell proliferation in acute myeloid leukemia cells (AML193 cell line) using MTT and tritiatedthymidine assays. They found that treatment of AML193 cells with Zebularine concentrations necessary to demethylate DNA causes a decrease in cell viability and proliferation, and an increase in the percentage of apoptotic cells and cells arrested in G₂/M of the cell cycle.

Calvisi et al. [17] reported that treatment with the demethylating agent Zebularine caused marked activation of MST1, p38MAPK, and JNK in hepatoma cell lines. Neureiter et al. [14] found that higher concentrations of Zebularine led to rapid induction of apoptosis in YAP C, DAN G, and Panc-89 pancreatic adenocarcinoma cells; lower concentrations remained ineffective in these cell lines. Billam et al. [18] found that Zebularine inhibits human breast cancer cell growth in a dose- and time-dependent manner.

Pharmacologic PCO prevention represents a valuable option, especially for refractive and pediatric cataract surgery. The current therapeutic concepts for pharmacologic PCO

prevention are compromised in terms of potential clinical application. The effective agents, represented by mitomycin-C [19] and 5-fluorouracil [20], require a sealed-capsule irrigation device due to significant toxic side effects [21]. Methylation inhibitor is a new strategy for treating disease. Methylation inhibitor is well tolerated and safe, according to a recent phase I clinical studies [22-24]. Further phase II clinical trials are undergoing. Other DNA methylation inhibitors, such as 5-Aza-C and 5-Aza-dC, have shown significant clinical activity against tumors. However, the clinical use of these agents is complicated by their toxicity and instability in aqueous solutions [13]. The half-life of Zebularine is much longer, and its toxicity is much lower. Therefore, Zebularine offers a better option for potential clinical application.

We have shown that Zebularine suppressed TGF β 2-induced lens epithelial cell-myofibroblast transdifferentiation [7]. In this study, our data demonstrate that Zebularine is an effective demethylating agent. Zebularine can inhibit LECs proliferation, attachment, and

migration in vitro. These findings suggest that Zebularine may become a new option for preventing PCO after conventional cataract surgery (Figure 5).

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REFERENCES

- Kohnen T, Fabian E, Gerl R, Hunold W, Hutz W, Strobel J, Hoyer H, Mester U. Optic edge design as long-term factor for posterior capsular opacification rates. *Ophthalmology* 2008; 115:1308-14. [PMID: 18321585]
- Jiang YX, Lu Y, Liu TJ, Yang J, Chen Y, Fang YW. Using HSV-TK/GCV suicide gene therapy to inhibit lens epithelial cell proliferation for treatment of posterior capsular opacification. *Mol Vis* 2011; 17:291-9. [PMID: 21283526]
- Awasthi N, Guo S, Wagner BJ. Posterior capsular opacification: a problem reduced but not yet eradicated. *Arch Ophthalmol* 2009; 127:555-62. [PMID: 19365040]
- Chan E, Mahroo OA, Spalton DJ. Complications of cataract surgery. *Clin Exp Optom* 2010; 93:379-89. [PMID: 20735786]
- Bhagwandien AC, Cheng YY, Wolfs RC, van Meurs JC, Luyten GP. Relationship between retinal detachment and biometry in 4262 cataractous eyes. *Ophthalmology* 2006; 113:643-9. [PMID: 16527355]
- Konno K, Nagamoto T. Membranous proliferation on the posterior surface of an intraocular lens after Nd:YAG laser capsulotomy. *Jpn J Ophthalmol* 2005; 49:173-5. [PMID: 15838738]
- Zhou P, Lu Y, Sun XH. Zebularine suppresses TGF-beta-induced lens epithelial cell-myofibroblast transdifferentiation by inhibiting MeCP2. *Mol Vis* 2011; 17:2717-23. [PMID: 22065925]
- Dwivedi RS, Herman JG, McCaffrey TA, Raj DS. Beyond genetics: epigenetic code in chronic kidney disease. *Kidney Int* 2011; 79:23-32. [PMID: 20881938]
- Hurd PJ, Whitmarsh AJ, Baldwin GS, Kelly SM, Waltho JP, Price NC, Connolly BA, Hornby DP. Mechanism-based inhibition of C5-cytosine DNA methyltransferases by 2-H pyrimidinone. *J Mol Biol* 1999; 286:389-401. [PMID: 9973559]
- Holleran JL, Parise RA, Joseph E, Eiseman JL, Covey JM, Glaze ER, Lyubimov AV, Chen YF, D'Argenio DZ, Egorin MJ. Plasma pharmacokinetics, oral bioavailability, and interspecies scaling of the DNA methyltransferase inhibitor, zebularine. *Clin Cancer Res* 2005; 11:3862-8. [PMID: 15897587]
- Beumer JH, Eiseman JL, Parise RA, Joseph E, Holleran JL, Covey JM, Egorin MJ. Pharmacokinetics, metabolism, and oral bioavailability of the DNA methyltransferase inhibitor 5-fluoro-2'-deoxycytidine in mice. *Clin Cancer Res* 2006; 12:7483-91. [PMID: 17138702]
- Cheng JC, Matsen CB, Gonzales FA, Ye W, Greer S, Marquez VE, Jones PA, Selker EU. Inhibition of DNA methylation and reactivation of silenced genes by zebularine. *J Natl Cancer Inst* 2003; 95:399-409. [PMID: 12618505]
- Scott SA, Lakshimikuttysamma A, Sheridan DP, Sanche SE, Geyer CR, DeCoteau JF. Zebularine inhibits human acute myeloid leukemia cell growth in vitro in association with p15INK4B demethylation and reexpression. *Exp Hematol* 2007; 35:263-73. [PMID: 17258075]
- Neureiter D, Zopf S, Leu T, Dietze O, Hauser-Kronberger C, Hahn EG, Herold C, Ocker M. Apoptosis, proliferation and differentiation patterns are influenced by Zebularine and SAHA in pancreatic cancer models. *Scand J Gastroenterol* 2007; 42:103-16. [PMID: 17190770]
- Zhou P, Zhao MW, Li XX, Yu WZ, Bian ZM. siRNA targeting mammalian target of rapamycin (mTOR) attenuates experimental proliferative vitreoretinopathy. *Curr Eye Res* 2007; 32:973-84. [PMID: 18027173]
- Hellebrekers DM, Jair KW, Vire E, Eguchi S, Hoebbers NT, Fraga MF, Esteller M, Fuks F, Baylin SB, van Engeland M, Griffioen AW. Angiostatic activity of DNA methyltransferase inhibitors. *Mol Cancer Ther* 2006; 5:467-75. [PMID: 16505122]
- Calvisi DF, Ladu S, Gorden A, Farina M, Conner EA, Lee JS, Factor VM, Thorgeirsson SS. Ubiquitous activation of Ras and Jak/Stat pathways in human HCC. *Gastroenterology* 2006; 130:1117-28. [PMID: 16618406]
- Billam M, Sobolewski MD, Davidson NE. Effects of a novel DNA methyltransferase inhibitor zebularine on human breast cancer cells. *Breast Cancer Res Treat* 2010; 120:581-92. [PMID: 19459041]
- Kim SY, Kim JH, Choi JS, Joo CK. Comparison of posterior capsule opacification in rabbits receiving either mitomycin-C or distilled water for sealed-capsule irrigation during cataract surgery. *Clin Experiment Ophthalmol* 2007; 35:755-8. [PMID: 17997781]
- Abdelwahab MT, Lindell I, Hultenby K, Kugelberg M. Transmission electron microscopy of the rabbit posterior capsule irrigated with thapsigargin and 5-fluorouracil in a sealed-capsule irrigation device. *Eye (Lond)* 2009; 23:975-8. [PMID: 18617905]
- Eibl KH, Liegl R, Kernt M, Priglinger S, Kampik A. Alkylphosphocholines as a potential pharmacologic prophylaxis for posterior capsule opacification. *J Cataract Refract Surg* 2009; 35:900-5. [PMID: 19393891]
- Quintas-Cardama A, Tong W, Kantarjian H, Thomas D, Ravandi F, Kornblau S, Manshouri T, Cortes JE, Garcia-Manero G, Verstovsek S. A phase II study of 5-azacitidine for patients with primary and post-essential thrombocythemia/polycythemia vera myelofibrosis. *Leukemia* 2008; 22:965-70. [PMID: 18385750]
- Braiteh F, Soriano AO, Garcia-Manero G, Hong D, Johnson MM, Silva Lde P, Yang H, Alexander S, Wolff J, Kurzrock R. Phase I study of epigenetic modulation with 5-azacytidine and valproic acid in patients with advanced cancers. *Clin Cancer Res* 2008; 14:6296-301. [PMID: 18829512]
- Plummer R, Vidal L, Griffin M, Lesley M, de Bono J, Coulthard S, Sludden J, Siu LL, Chen EX, Oza AM, Reid GK, McLeod AR, Besterman JM, Lee C, Judson I, Calvert H, Boddy AV. Phase I study of MG98, an oligonucleotide antisense inhibitor of human DNA methyltransferase 1, given as a 7-day infusion in patients with advanced solid tumors. *Clin Cancer Res* 2009; 15:3177-83. [PMID: 19383817]

