

# The development of meibomian glands in mice

Chyong Jy Nien,<sup>1</sup> Salina Massei,<sup>1</sup> Gloria Lin,<sup>1</sup> Hongshan Liu,<sup>2</sup> Jerry R. Paugh,<sup>3</sup> Chia-Yang Liu,<sup>2</sup> Winston Whei-Yang Kao,<sup>2</sup> Donald J. Brown,<sup>1</sup> James V. Jester<sup>1</sup>

<sup>1</sup>Gavin Herbert Eye Institute, University of California Irvine, CA; <sup>2</sup>Ophthalmology Department, University of Cincinnati, OH;

<sup>3</sup>Southern California College of Optometry, Fullerton, CA

**Purpose:** The purpose of this study was to characterize the natural history of meibomian gland morphogenesis in the mouse.

**Methods:** Embryonic (E) and post natal (P) C57Bl/6 mouse pups were obtained at E18.5, P0, P1, P3, P5, P8, P15, and P60. Eyelids were fixed and processed for en bloc staining with Phalloidin/DAPI to identify gland morphogenesis, or frozen for immunohistochemistry staining with Oil red O (ORO) to identify lipid and antibodies specific against peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ) to identify meibocyte differentiation. Samples were then evaluated using a Zeiss 510 Meta laser scanning confocal microscope or Nikon epi-fluorescent microscope. Tissues from adult mice (2 month-old) were also collected for western blotting.

**Results:** Meibomian gland morphogenesis was first detected at E18.5 with the formation of an epithelial placode within the fused eyelid margin. Invagination of the epithelium into the eyelid was detected at P0. From P1 to P3 there was continued extension of the epithelium into the eyelid. ORO and PPAR $\gamma$  staining was first detected at P3, localized to the central core of the epithelial cord thus forming the presumptive ductal lumen. Ductal branching was first detected at P5 associated with acinar differentiation identified by ORO and PPAR $\gamma$  staining. Adult meibomian glands were observed by P15. Western blotting of meibomian gland proteins identified a 50 kDa and a 72 kDa band that stained with antibodies specific to PPAR $\gamma$ .

**Conclusions:** We have demonstrated that meibomian gland development bears distinct similarities to hair development with the formation of an epithelial placode and expression of PPAR $\gamma$  co-incident with lipid synthesis and meibocyte differentiation.

Meibomian glands are greatly enlarged modified sebaceous glands without a hair follicle [1] and as a skin appendage, their development is thought to share similarity with hair and sebaceous gland morphogenesis in the skin. During development of pilosebaceous units, mesenchymal-epithelial interactions at embryonic day 14.5 (E14.5) lead to the formation of an ectodermal-epithelial placode that progressively undergoes proliferation and growth to form hair follicles that break the skin surface after birth [2]. Sebaceous gland precursors appear to form along the upper segment of the hair follicle toward the end of embryogenesis and are characterized by the expression of peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ) and the synthesis of lipid [3]. PPARs play critical roles in cell proliferation, differentiation, and apoptosis [4]. They are implicated in fetal maturation of murine epidermis, regulating keratinocyte differentiation, and the morphogenesis of skin appendages including hair follicles and sebaceous glands. The PPAR $\gamma$  isoform is the most studied receptor and is thought to be a key regulator of adipogenesis and adipocyte differentiation [5,6]. PPAR $\gamma$  is also required for the differentiation of the sebaceous

gland [7] and has been shown to be present in mouse and human skin sebaceous glands in vivo [8].

By comparison, little is known about meibomian gland development [9,10] and the most extensive study, reported by Andersen, et al. [11], focuses on the human meibomian gland. More importantly, no studies have reported on the expression of PPAR $\gamma$  during meibomian gland development and its association with lipidogenesis.

Recently, a role for PPAR $\gamma$  in meibomian gland function was hypothesized based on findings that PPAR $\gamma$  localization undergoes specific age-related changes in older mouse meibomian glands [12]. In young mice (<6 months of age), PPAR $\gamma$  is localized in small vesicles within the cytoplasm and the nucleus that is associated with a high meibocyte proliferation index. Aging mice (>1 year of age) show primarily a nuclear localization that is associated with decreased meibocyte proliferation and meibomian gland atrophy. Overall, these findings suggest that altered PPAR $\gamma$  receptor signaling in older mice may control changes in cell cycle entry/proliferation, lipid synthesis and gland atrophy during aging that may underlie age-related meibomian gland dysfunction.

In this report we provide evidence that meibomian gland development bears distinct similarities to sebaceous gland development and that PPAR $\gamma$  expression coincides with

---

Correspondence to: James V. Jester, Ph.D., Gavin Herbert Eye Institute, University of California Irvine Medical Center, 101 The City Drive, Orange, CA, 92868; Phone: (714) 456-5022; FAX: (714) 456-5073; email: [jjester@uci.edu](mailto:jjester@uci.edu)

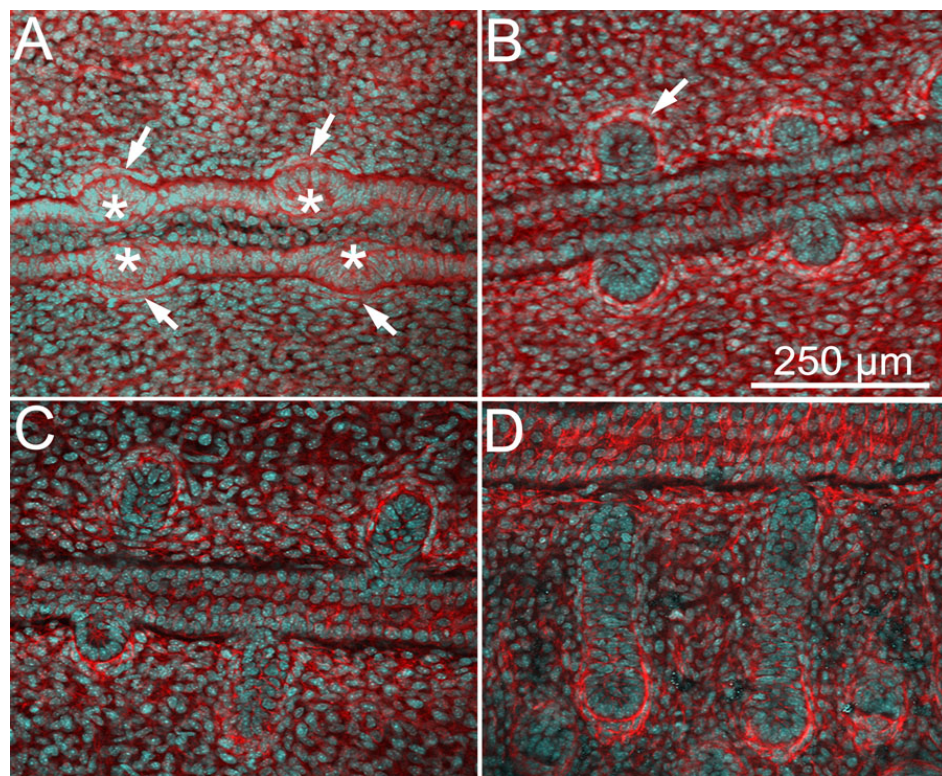


Figure 1. Confocal images using actin (red) and nuclei (DAPI, cyan) staining of eyelids at different time points of meibomian gland morphogenesis. At E18.5 (A), we observed the formation of an epithelial condensation (asterisk) within the fused lid margin. Epithelial placodes in the superior and inferior lids also appeared opposite to each other, and were associated with condensation of mesenchyme as detected by cell alignment and increased actin staining of mesenchyme directly adjacent to the placode (arrows). At P0 (B), we observed an epidermal invagination (arrow) that undergoes proliferation and maturation with progressive enlargement and elongation adopting a tubular shape at P1 and P3 (C and D, respectively).

lipogenesis and meibocyte differentiation. Overall these findings support the hypothesis that PPAR $\gamma$  is a major regulator of meibomian gland function.

## METHODS

**Animals:** C57Bl/6 mice were used in this study. Embryonic (E) and postnatal (P) mouse pups were obtained at E18.5, P0, P1, P3, P5, P8, P15, and P60. Four mice were sacrificed at each time point.

**Confocal microscopy:** Eyelids from E18.5, P0, P3, P5, P8, and P15 were dissected and fixed in 4% paraformaldehyde overnight at 4 °C. Samples were then washed 2 times for 30 min each with 50% TD buffer (0.5% DMSO, 0.5% Triton X, 2.5% Dextran 40 in PBS, PH 7.4) at 4 °C. Tissue was then stained en bloc with Rhodamine Phalloidin (dilution 1:20 in 50% TD buffer; Molecular Probes, Invitrogen, Carlsbad, CA) and DAPI (4',6-diamidino-2-phenylindole nuclei stain, dilution 1:1,000; Molecular Probes, Invitrogen, Carlsbad, CA) overnight on a rocker at 4 °C. The next day, samples were washed with 50% TD buffer, 3 times, 30 min each, transferred to PBS, and scanned using a confocal microscope (LSM 510; Carl Zeiss MicroImaging, Thornwood, NY) using 543 nm and 633 nm lasers under a 40 $\times$  oil objective.

**Immunohistochemistry:** Eyelids were fixed in 2% PFA overnight, placed in 15% sucrose for 4 h and then transferred to 30% sucrose overnight. Lids were embedded in Tissue-Tek® embedding medium for frozen tissue (Sakura Finetek USA, Inc, Torrance, CA), frozen in

liquid nitrogen and blocks stored at -80 °C until sectioned on a cryostat. Cryosections (8 μm thick) were then cut mounted on glass slides, and either stained for Oil Red O (ORO, Sigma-Aldrich, St. Louis, MO) or with antibodies to PPAR $\gamma$ . Sections were stained with ORO by first rinsing in PBS and then incubating in 0.5% ORO solution for 10 min. Sections were then rinsed with PBS and counterstained with hematoxylin.

For immunostaining, tissue sections were rehydrated with PBS, placed in acetone for 3 min and then re-hydrated in PBS. Sections were then blocked with 1% BSA in PBS for 30 min at 37 °C and then stained with rabbit anti-PPAR $\gamma$  (1:50; Abcam, Cambridge, MA) for 1 h at 37 °C. Sections were then washed with PBS, stained with FITC conjugated goat anti-rabbit IgG (dilution 1:200; Invitrogen, Carlsbad, CA) for 1 h at 37 °C and then counterstained with DAPI. For negative controls, primary antibodies were omitted. The samples were then evaluated and imaged using a Nikon Eclipse E600 fluorescence microscope.

**Western blotting:** Eyelids from P60 mice were removed and meibomian glands dissected from both the upper and lower eye lids. Tissue was weighed and 39 mg of meibomian glands from 5 mice were then placed in 0.5 ml of Radioimmunoprecipitation assay buffer (RIPA buffer, 25 mM Tris-HCl pH 7.6, 150 mM NaCl, 1% NP-40, 1% sodium deoxycholate, 0.1% SDS) containing protease inhibitor and phosphatase inhibitor. Tissue was centrifuged at 5,000 $\times$  g for 20 min and the supernatant was collected. Total protein was then measured using the RC DC Protein Assay (Bio-Rad

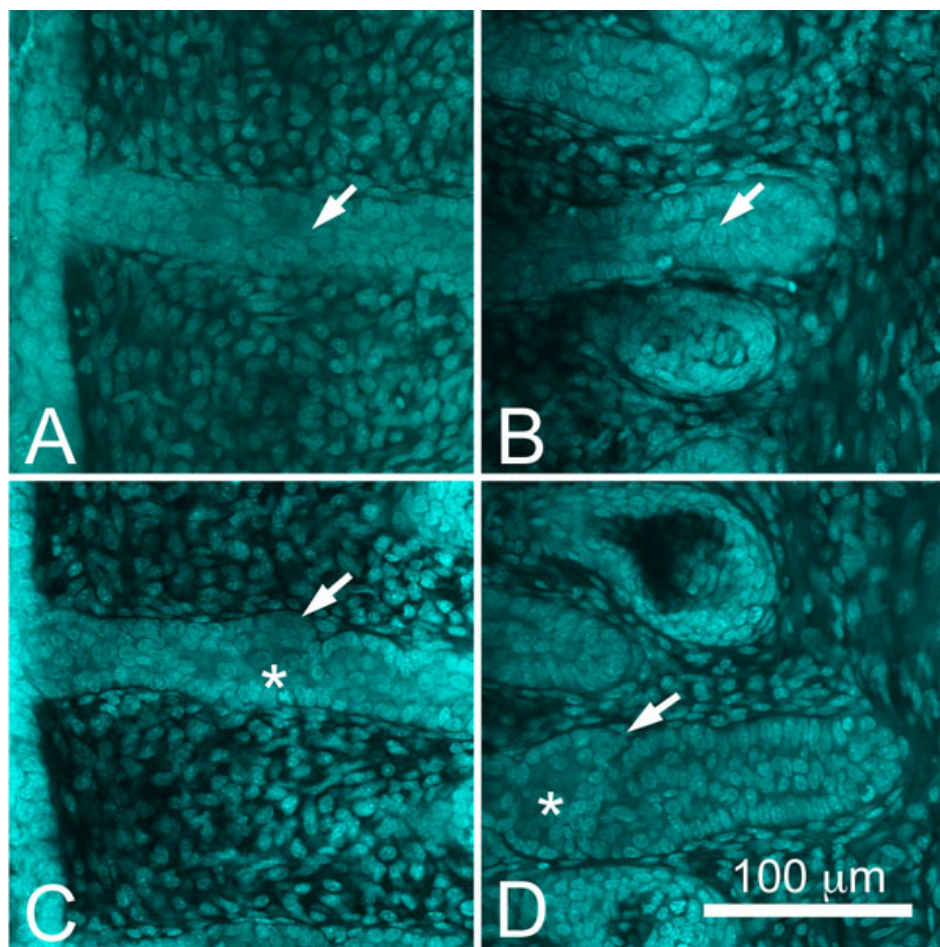


Figure 2. Nuclei staining (DAPI) at P3 and P5. At P3 (**A** and **B**), meibomian glands appeared to be comprised of a cord of epithelium with a bordering basal cell layer and a two-three cell suprabasal cell layer within the inner core at the distal end of the developing meibomian gland (**A**, arrow). At the proximal end there appeared to be a reduction of the number of cells within the inner core of the developing duct (**B**, arrow). By P5 (**C** and **D**), early branching of the invaginating epithelial cord was observed (arrows). Additionally, there appeared to be enlargement of cells within the central cord of the epithelium (asterisk).

Laboratories, Hercules, CA). For comparison, samples from white and brown fat were also obtained and protein collected. White and brown fat tissue was frozen in liquid nitrogen and placed in 1 ml of RIPA buffer containing protease inhibitor and phosphatase inhibitor. The tissue samples were sonicated, then centrifuged at low speed  $1,500\times g$  for 5 min at  $4\text{ }^{\circ}\text{C}$  to obtain the supernatant. The supernatant was centrifuged again at  $4\text{ }^{\circ}\text{C}$  under the “Fast Cool” function of the centrifuge. Samples were run on 10% SDS PAGE and the proteins were transferred to PVDF membrane via iBlot Gel Transfer Device (Invitrogen). The blot was then blocked using 5% non-fat dry milk in TBS-T (0.2% Tween-20). Following blocking, the blot was immunostained using antibodies to PPAR $\gamma$  (1:500 Dilution) and rinsed with TBS-T (0.2% Tween-20). The blot was then incubated with goat anti-rabbit IgG (H+L) HRP (Dilution 1:2,500; Thermo Scientific, Rockford, IL) and rinsed with TBS-T (0.2% Tween-20). To visualize the bands, the blot was incubated with SuperSignal West Pico Chemiluminescent Substrate (Thermo Scientific, Rockford, IL) and detected using photographic film.

## RESULTS

*Meibomian gland morphogenesis:* Meibomian gland development appeared to start at E18.5 (Figure 1A) with the formation of an epithelial placode (Figure 1A; asterisk) within the fused eyelid margin epithelium. Epithelial placodes in the superior and inferior lids also appeared to form in tandem, immediately opposite to each other, and were associated with condensation of mesenchyme as detected by cell alignment and increased actin staining of mesenchyme directly adjacent to the placode (Figure 1A; arrows). At birth (P0, Figure 1B), the epithelial invagination into the eyelid mesenchyme was first detected and appeared to form a solid cord with no clear lumen (Figure 1B, arrow) and with more prominent condensation of mesenchyme surrounding the epithelial invagination. Progressive lengthening of the invaginating epithelial cord was detected from P1 (Figure 1C) to P3 (Figure 1D) with little additional morphogenetic changes.

At higher magnification, developing meibomian glands at P3 appeared to be comprised of a cord of epithelium with a bordering basal cell layer and a two-three cell suprabasal cell layer within the inner core at the distal end of the developing meibomian gland (Figure 2A, arrow). At the proximal end of the developing gland there appeared to be a

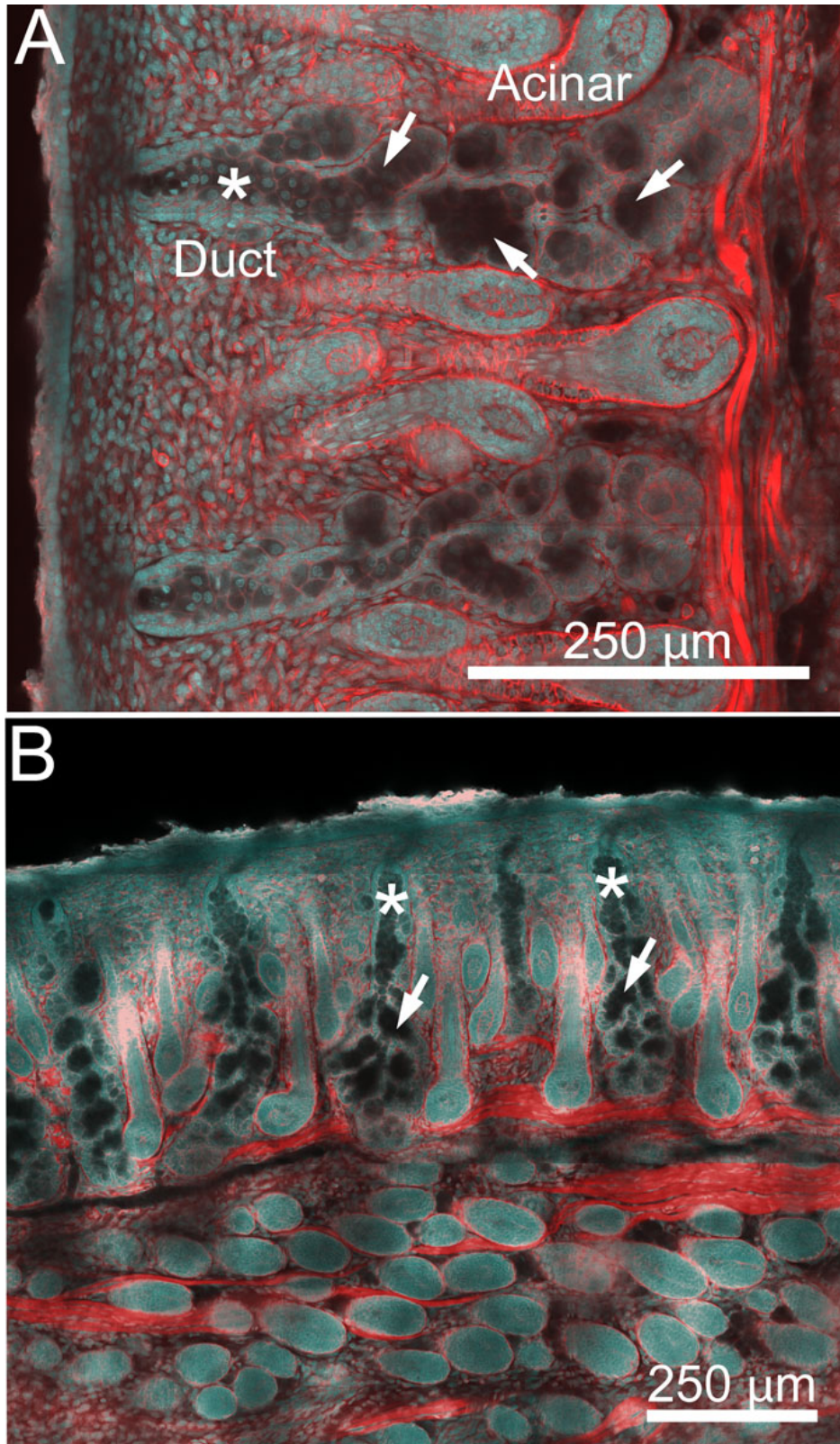


Figure 3. Confocal images using actin (red) and nuclei (DAPI, cyan) staining of eyelids at P8 and P15. At P8 (A), developing meibomian glands showed distinct differentiation into ductal and acinar regions. Within the ductal region, basal epithelial cells along with a single layer of suprabasal cells appear to line a more central region containing enlarged epithelial cells that lacked actin staining (asterisk). Within the acinar region, the developing meibomian gland duct branched into multiple ductules that appeared to lack any cellular contents (arrows) and terminated into developing acini. At P15 (B), the meibomian glands appear to obtain an adult meibomian gland morphology.

reduction of the number of cells within the inner core of the developing duct (Figure 2B, arrow). By P5, early branching

of the invaginating epithelial cord was detected at both the distal (Figure 2C, arrow) and proximal end (Figure 2D, arrow)

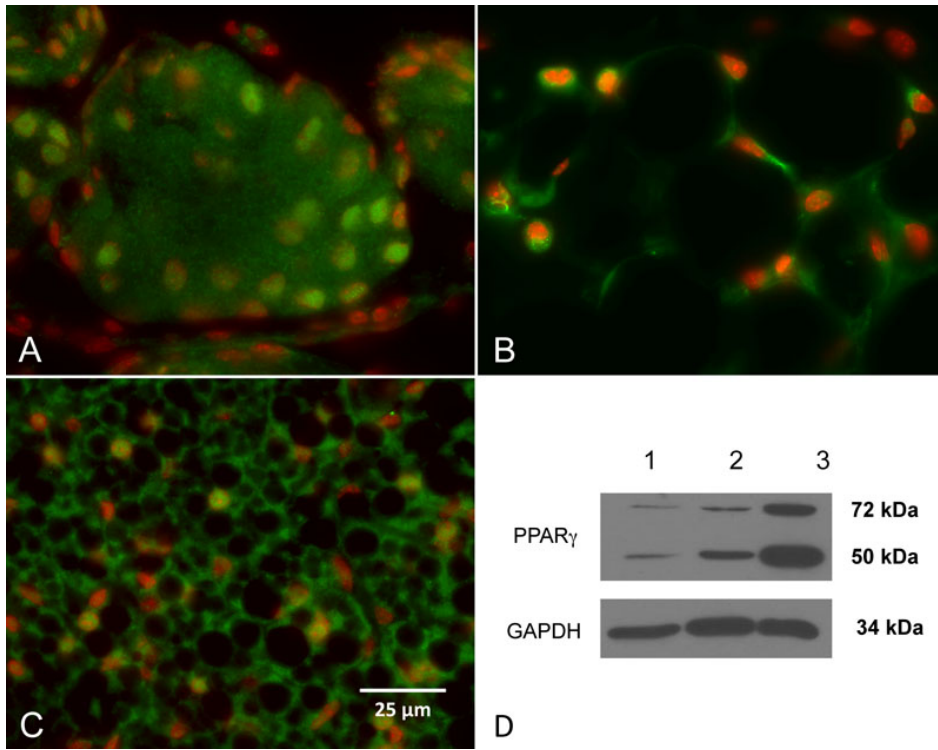


Figure 4. PPAR $\gamma$  (Green) and nuclei (DAPI, red) staining of meibomian glands, white fat, and brown fat from adult, P60 mice. In the meibomian glands (A), nuclear and extra-nuclear staining of the plasma membrane and intracellular structures, possibly microsomes were detected. Staining of white fat and brown fat (B and C, respectively) showed predominantly nuclear staining. Western blotting of meibomian gland protein with PPAR $\gamma$  antibodies identified a 50 kDa protein band (D, lane 1) that was also present in extracts from white fat and brown fat (D, lane 2 and 3, respectively). Note that the meibomian gland, white fat and brown fat all showed a second band with a molecular weight of 72 kDa that may represent post-translational modifications of PPAR $\gamma$ .

of the developing gland. Additionally, there appeared to be enlargement of cells within the central cord of the epithelium (Figure 2C,D, asterisk). By P8 the developing meibomian glands showed distinct differentiation into ductal and acinar regions (Figure 3A). Within the ductal region, basal epithelial cells along with a single layer of suprabasal cells appeared to line a more central region containing enlarged epithelial cells that lacked actin staining (Figure 3A, asterisk). Within the acinar region, the developing meibomian gland duct branched into multiple ductules that appeared to lack any cellular contents (arrows) and terminated into developing acini. By P15, the meibomian glands appeared to obtain a normal meibomian gland morphology (Figure 3B).

*Meibocyte differentiation and lipid synthesis:* Meibocyte differentiation was evaluated by staining for PPAR $\gamma$  using a rabbit polyclonal antibody directed against a synthetic peptide derived from residues 50–150 of human PPAR $\gamma$ . Immunostaining of meibomian glands from adult, P60 mice showed nuclear and extra-nuclear staining of the plasma membrane and intracellular structures, possibly microsomes (Figure 4A). Staining of white fat and brown fat (Figure 4B,C, respectively) showed predominantly nuclear staining. Western blotting of meibomian gland protein from adult mice with PPAR $\gamma$  antibodies identified a 50 kDa protein band (Figure 4D, lane 1) that was also present in extracts from white fat and brown fat (Figure 4D, lane 2 and 3, respectively). An additional band at 72 kDa was also identified that might be related to post-translational modifications of the protein.

During morphogenesis, staining for PPAR $\gamma$  was negative at E18.5 and at birth (P0) (Figure 5A,B, respectively). PPAR $\gamma$  staining was first detected at P3 within the central region of the invaginating epithelium (Figure 5C, arrow); the same time that sebaceous glands appeared to stain for PPAR $\gamma$  (arrowhead). PPAR $\gamma$  staining appeared to progressively increase at P5 and P8 as the gland continued to develop (Figure 5D,E, arrow), however the localization appeared to be both within and along the proximal end of developing meibomian gland duct. By P15, PPAR $\gamma$  staining was localized to the meibomian gland acini (Figure 5F, arrows) that bordered the central ductal epithelium, which was negative for PPAR $\gamma$  (Figure 5F, asterisk).

Association of meibocyte differentiation with lipid production was assessed by ORO staining and showed that eyelids were negative for lipid at both E18.5 and P0 (Figure 6A,B, arrow). Lipid was first detected at P3 and was limited to the central region of the invaginating epithelium (Figure 6C, arrow); the same region stained by antibodies to PPAR $\gamma$ . At P5 and P8, ORO staining continued to be limited to the inner core of the developing duct and ductules (Figure 6D,E, respectively). After eyelid opening at P15, ORO was present in the acini of the mature meibomian glands (Figure 6F).

## DISCUSSION

To our knowledge, reports covering the development of the meibomian gland are fragmentary and limited, for the most part, to the appearance of tarsal glands during eyelid opening

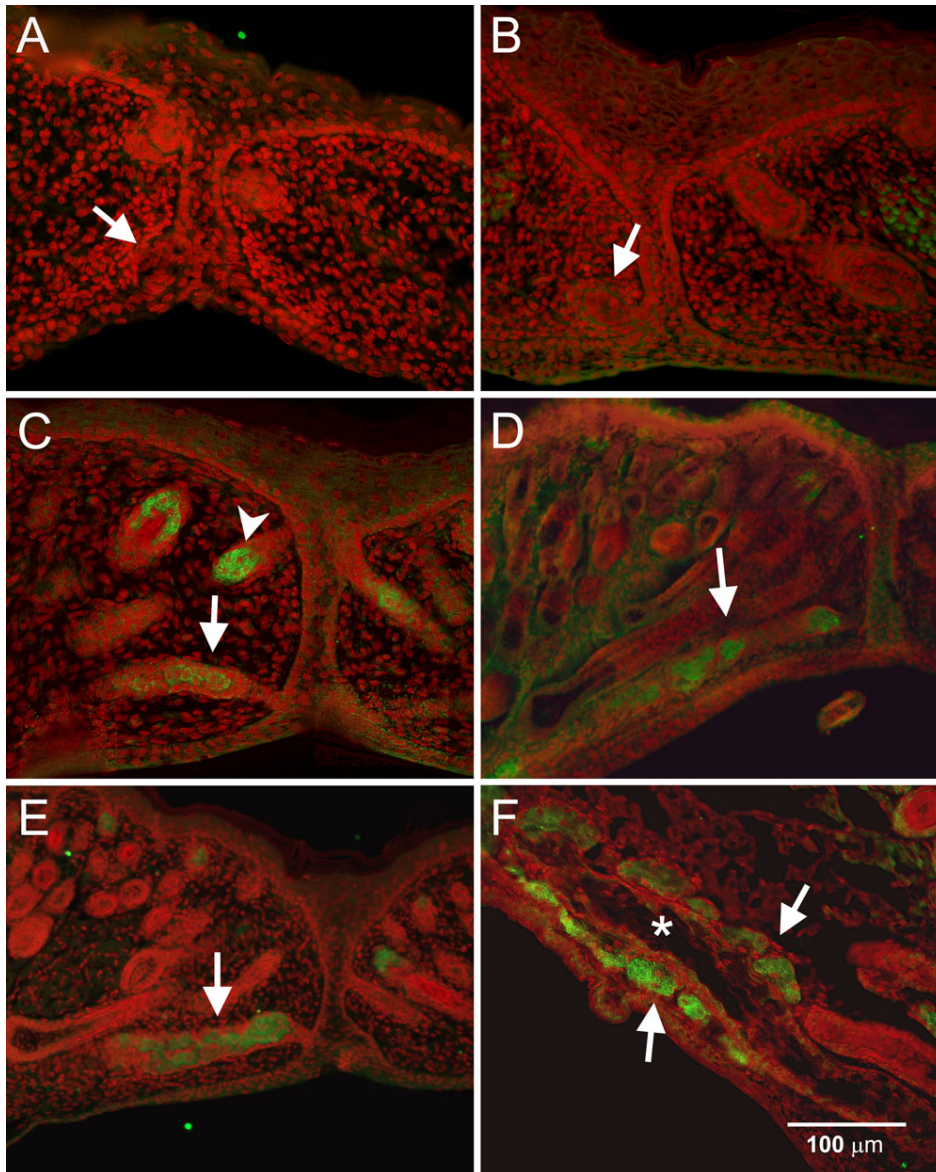


Figure 5. PPAR $\gamma$  (green) and nuclei (DAPI, red) staining during meibomian gland morphogenesis. At E18.5 (A) and P0 (B), PPAR $\gamma$  staining of the epithelial placode was negative. At P3 (C), PPAR $\gamma$  was first detected in the lumen of the developing gland and increased progressively as the gland continued to develop at P5 (D) and P8 (E). At P15 (F), PPAR $\gamma$  staining was limited to the acini (arrows) or the terminal regions of the branching ductules (the asterisk shows a well developed central duct at P15).

[9,10]. The most extensive study reported by Andersen et al. [11] and also discussed by Knop et al. [13] reports that meibomian glands in the human eyelid initially form as epithelial cords that develop following eyelid fusion. Lateral branching of the epithelial cords then form acini later in development. Lipid synthesis was also noted to occur within the acini and the epithelial invagination thus forming the central ductal system of the glands.

Our results with confocal microscopy support and extend these observations and indicate that meibomian gland development in the mouse is initiated around E18.5 with the formation of an epithelial placode and mesenchymal condensation, similar to that observed in hair follicle development [14,15]. Invagination of the epithelium into the developing mesenchyme then continues from birth to about

P3 with initial branching of the epithelial cord detected at P5. By P8 the developing meibomian gland shows extensive ductal branching and the formation of distinct acini with mature meibomian glands present by P15 or eyelid opening.

During meibomian gland development, PPAR $\gamma$  expression was first detected at P3, before epithelial branching and the formation of distinct acini. PPAR $\gamma$  expression was also localized to the central core of the invaginating epithelium, the same region where lipid synthesis was first detected. Interestingly, PPAR $\gamma$  expression remained localized to the developing duct and acinar regions from P5 to P8, but was restricted to the acini by P15. Overall, these findings suggest that PPAR $\gamma$  plays an important role in meibomian gland morphogenesis, meibocyte differentiation and lipid synthesis.

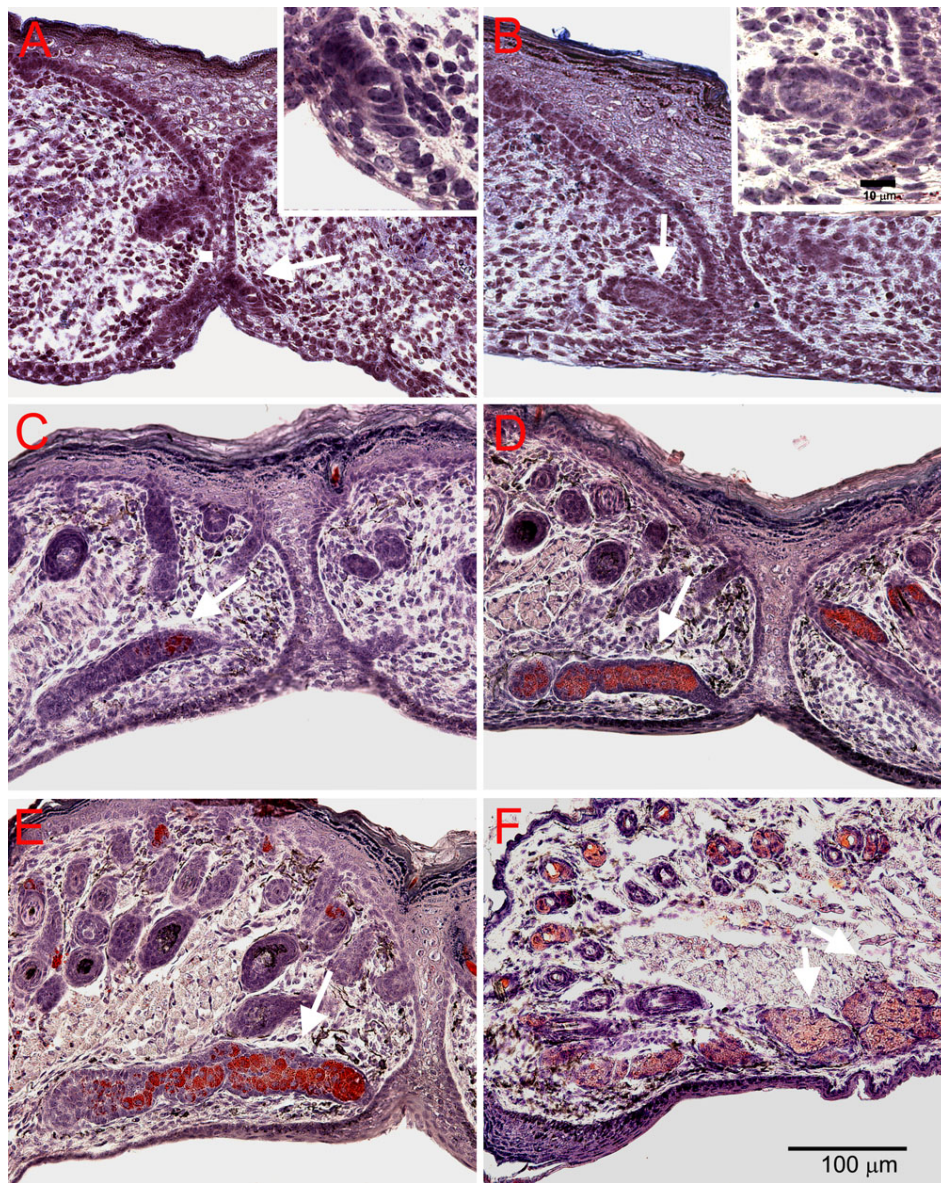


Figure 6. ORO staining for lipid during meibomian gland development at different time points. Consistent with PPAR $\gamma$ , ORO staining was absent at E18.5 (A) and P0 (B). Lipid production was first detected at P3 (C) within the central region of the developing duct. At P5 (D) and P8 (E), ORO staining continued to be limited to the inner core of the developing duct. After eyelid opening at P15 (F), ORO was present in the acini of the mature gland.

Interestingly, western blots of adult meibomian gland tissue extracts identified a low and high molecular weight PPAR $\gamma$  protein. Message for PPAR $\gamma$  can be alternatively spliced to a 475 amino acid ( $\gamma$ 1) and 505 amino acid ( $\gamma$ 2) isoform of approximately 50 kDa. PPAR $\gamma$  also contains a consensus serine phosphorylation site at S112 that may be associated with decreased transcriptional activity [16,17] and two sumoylation sites, K107 and K395, that may be linked to nuclear-cytoplasmic transport, apoptosis, and transcriptional regulation [18]. Since sumoylation covalently attaches sumo-1 or sumo-2/3, each approximately 11 kDa, it is likely that the high molecular weight PPAR $\gamma$  identified in this study represents a post translational sumoylation. Clearly, future studies identifying the post translational modifications of

PPAR $\gamma$  will be important to understanding the role of this receptor in regulating meibomian gland function.

Additionally, PPAR $\gamma$  contains an NH $_2$ -terminal transactivation domain (residues 30–136), a DNA binding domain (residues 136–140), and a COOH-terminal ligand-binding domain (residues 204–505) containing a ligand-dependent transactivation function [19]. Interestingly, PPAR $\gamma$  functions as a “molecular switch” and can recruit corepressors, N-CoR and SMRT, when unliganded PPAR $\gamma$  binds to some target genes promoters, which upon ligand binding recruits co-activators, SRC1/CBP and TRAP/DRIP/ARC complexes [20].

Importantly, PPAR $\gamma$  has been shown to play a critical role in both adipogenesis and sebaceous gland development with mice chimeric for PPAR $\gamma$  null cells showing little or no

contribution to the formation of fat or sebaceous glands [7]. Additionally, PPAR $\gamma$  agonists, such as thiazolidinediones and rosiglitazone, increase sebum production in diabetic patients and lipid synthesis in sebocyte cell cultures, respectively [21]. Recent studies also suggest that there can be distinct gender differences in the transcriptional activity of the *PPAR* gene [22] and there is at least one report suggesting that androgens and PPAR ligands may act synergistically to regulate lipid synthesis in cultured sebocytes [23]. If transferable to the meibomian gland, these findings suggest that understanding the role of PPAR $\gamma$  in meibomian gland function may provide important insights into the pathogenesis of meibomian gland dysfunction. Further work studying the modulation of PPAR $\gamma$  expression or activity will be needed to clarify the role PPAR $\gamma$  in regulating meibum production. More interestingly, selective modulation of PPAR $\gamma$  activity may be a potential therapeutic strategy for the treatment of dry eye syndrome and meibomian gland dysfunction.

In summary, we have demonstrated that meibomian gland development starts with the formation of an epithelial placode similar to that observed for the development of the pilosebaceous unit. Development then proceeds with invagination of the epithelium into the eyelid mesenchyme forming a solid cord. By post natal day 3, the adipogenic nuclear receptor PPAR $\gamma$  is first expressed within the epithelial cord in association with lipid synthesis, thus forming the presumptive ductal lumen, and later, the nascent acini at post natal day 5. Meibomian glands appear to be completely formed by the time of eyelid opening with PPAR $\gamma$  restricted to the acinar compartment. Overall, PPAR $\gamma$  appears to be a molecular marker of meibocyte differentiation.

#### ACKNOWLEDGMENTS

Supported by NEI EY016663, Discovery Eye Foundation, The Skirball Program in Molecular Ophthalmology, Research to Prevent Blindness, and an unrestricted grant from Alcon Research.

#### REFERENCES

1. Kozak I, Bron AJ, Kucharova K, Kluchova D, Marsala M, Heichel CW, Tiffany JM. Morphologic and volumetric studies of the meibomian glands in elderly human eyelids. *Cornea* 2007; 26:610-4. [PMID: 17525661]
2. Fuchs E. Skin stem cells: rising to the surface. *J Cell Biol* 2008; 180:273-84. [PMID: 18209104]
3. Michalik L, Wahli W. Peroxisome proliferator-activated receptors (PPARs) in skin health, repair and disease. *Biochim Biophys Acta* 2007; 1771:991-8. [PMID: 17400022]
4. Keller JM, Collet P, Bianchi A, Huin C, Bouillaud-Kremarik P, Becuwe P, Schohn H, Domenjoud L, Dauca M. Implications of peroxisome proliferator-activated receptors (PPARs) in development, cell life status and disease. *Int J Dev Biol* 2000; 44:429-42. [PMID: 11032176]
5. Mao-Qiang M, Fowler AJ, Schmuth M, Lau P, Chang S, Brown BE, Moser AH, Michalik L, Desvergne B, Wahli W, Li M, Metzger D, Chambon PH, Elias PM, Feingold KR. Peroxisome-proliferator-activated receptor (PPAR)-gamma activation stimulates keratinocyte differentiation. *J Invest Dermatol* 2004; 123:305-12. [PMID: 15245430]
6. Sandouk T, Reda D, Hofmann C. The antidiabetic agent pioglitazone increases expression of glucose transporters in 3T3-F442A cells by increasing messenger ribonucleic acid transcript stability. *Endocrinology* 1993; 133:352-9. [PMID: 8319581]
7. Rosen ED, Sarraf P, Troy AE, Bradwin G, Moore K, Milstone DS, Spiegelman BM, Mortensen RM. PPAR gamma is required for the differentiation of adipose tissue in vivo and in vitro. *Mol Cell* 1999; 4:611-7. [PMID: 10549292]
8. Rosen ED, Spiegelman BM. PPARgamma: a nuclear regulator of metabolism, differentiation, and cell growth. *J Biol Chem* 2001; 276:37731-4. [PMID: 11459852]
9. Findlater GS, McDougall RD, Kaufman MH. Eyelid development, fusion and subsequent reopening in the mouse. *J Anat* 1993; 183:121-9. [PMID: 8270467]
10. Teraishi T, Yoshioka M. Electron-microscopic and immunohistochemical studies of eyelid reopening in the mouse. *Anat Embryol (Berl)* 2001; 204:101-7. [PMID: 11556525]
11. Andersen H, Ehlers N, Matthiessen ME. Histochemistry and development of the human eyelids. *Acta Ophthalmol (Copenh)* 1965; 43:642-68. [PMID: 4159407]
12. Nien CJ, Paugh JR, Massei S, Wahlert AJ, Kao WW, Jester JV. Age-related changes in the meibomian gland. *Exp Eye Res* 2009; 89:1021-7. [PMID: 19733559]
13. Knop N, Knop E. Meibomian glands. Part I: anatomy, embryology and histology of the Meibomian glands. *Ophthalmologe* 2009; 106:872-83. [PMID: 19856010]
14. Fuchs E. Scratching the surface of skin development. *Nature* 2007; 445:834-42. [PMID: 17314969]
15. Kao WW, Xia Y, Liu CY, Saika S. Signaling pathways in morphogenesis of cornea and eyelid. *Ocul Surf* 2008; 6:9-23. [PMID: 18264652]
16. Adams M, Reginato MJ, Shao D, Lazar MA, Chatterjee VK. Transcriptional activation by peroxisome proliferator-activated receptor gamma is inhibited by phosphorylation at a consensus mitogen-activated protein kinase site. *J Biol Chem* 1997; 272:5128-32. [PMID: 9030579]
17. Hu E, Kim JB, Sarraf P, Spiegelman BM. Inhibition of adipogenesis through MAP kinase-mediated phosphorylation of PPARgamma. *Science* 1996; 274:2100-3. [PMID: 8953045]
18. Geiss-Friedlander R, Melchior F. Concepts in sumoylation: a decade on. *Nat Rev Mol Cell Biol* 2007; 8:947-56. [PMID: 18000527]
19. Tontonoz P, Spiegelman BM. Fat and beyond: The diverse biology of PPARgamma. *Annu Rev Biochem* 2008; 77:289-312. [PMID: 18518822]
20. van Beekum O, Fleskens V, Kalkhoven E. Posttranslational modifications of PPAR-gamma: fine-tuning the metabolic master regulator. *Obesity (Silver Spring)* 2009; 17:213-9. [PMID: 19169221]
21. Trivedi NR, Cong Z, Nelson AM, Albert AJ, Rosamilia LL, Sivarajah S, Gilliland KL, Liu W, Mauger DT, Gabbay RA, Thiboutot DM. Peroxisome proliferator-activated receptors increase human sebum production. *J Invest Dermatol* 2006; 126:2002-9. [PMID: 16675962]

22. Ciana P, Biserni A, Tatangelo L, Tiveron C, Sciarroni AF, Ottobrini L, Maggi A. A novel peroxisome proliferator-activated receptor responsive element-luciferase reporter mouse reveals gender specificity of peroxisome proliferator-activated receptor activity in liver. *Mol Endocrinol* 2007; 21:388-400. [PMID: 17158222]
23. Makrantonaki E, Zouboulis CC. Testosterone metabolism to 5 $\alpha$ -dihydrotestosterone and synthesis of sebaceous lipids is regulated by the peroxisome proliferator-activated receptor ligand linoleic acid in human sebocytes. *Br J Dermatol* 2007; 156:428-32. [PMID: 17300229]